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Coccolithophore ecology in the tropical and subtropical Atlantic Ocean: New perspectives from the Atlantic meridional transect (AMT) programme

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ABSTRACT

Coccolithophore species composition was determined in 199 samples collected from the upper 300 m of the Atlantic Ocean, spanning temperate, tropical and subtropical waters in both hemispheres during four Atlantic Meridional Transect (AMT) cruises over the period 2003–2005. Of the 171 taxa observed, 140 consistently represented <5% of total cell numbers, and were classed as rare. Multivariate statistical techniques were used on the common taxa to assess variability in community composition vertically in the water column, horizontally across hydrographic provinces (subtropical gyres, equatorial waters, temperate waters), and temporally between cruises. Sharper gradients of statistical dissimilarity in species composition occurred vertically over a few tens of metres than horizontally over hundreds of kilometres. Three floral groups were identified from analysis of the depth of normalised abundance maxima in the subtropical gyres and equatorial waters: the upper euphotic zone (UEZ, >10% surface irradiance); the lower euphotic zone (LEZ, 10–1% surface irradiance); and the sub-euphotic zone (SEZ, <1% surface irradiance). The LEZ includes the deep chlorophyll maximum (DCM) and nutricline, and was characterised by species such as Emiliania huxleyi and Gephyrocapsa ericsonii which were also abundant at higher latitudes. It is suggested that this pattern reflects similarities in the light (and inorganic nutrient) conditions between the LEZ and temperate waters. The SEZ is below the depth where light is thought to be sufficient to support photosynthesis, suggesting that deep-dwelling species such as Florisphaera profunda and Gladiolithus spp. may be mixotrophic or phagotrophic, although conclusive proof will need to be gained experimentally. Mixotrophy could also be an important nutritional strategy for species abundant (Umbellosphaera spp., holococcolithophores) in the UEZ where inorganic nutrient concentrations are depleted and limiting to growth, although other nutritional strategies, such as the use of organic nutrients, are also possible. Statistical differences were also found in the species composition between the different cruises, with high levels of similarity for similar timed cruises (May or September–October). Few individual taxa showed significant variability in abundance over the time-span of sampling, except species such as E. huxleyi and G. ericsonii at higher latitudes. In subtropical and equatorial waters, high levels of species richness and low levels of species dominance remained throughout the sampling period indicating that seasonal fluctuations reflected differences in the whole coccolithophore community rather than in just one or a few species. Multivariate analyses of the taxa classified as rare also indicated some level of temporal, as well as vertical, zonation. Such insights into coccolithophore ecology and community composition provide important new perspectives that require innovative research to fully understand their impact on ocean biogeochemistry.

1. Introduction

Coccolithophores are unicellular marine algae, belonging to the Class Prymnesiophyceae, that possess one or more external layers of calcite scales (coccoliths). Coccoliths are formed intracellularly via calcification and are then extruded onto the external surface
of the cell to form a coccosphere. Around 200 species of coccolithophore are extant in the modern ocean (Young et al., 2003), with considerable diversity in the shape of the cell, the shape, construction and crystallography of the coccoliths, and their number, diversity and geometry around the cell (Monteiro et al., 2016). Variability in the crystallography of the coccoliths is based around two forms which represent linked species' life stages and reproductive strategies; in their diploid form many species produce heterococcoliths (HET), composed of complex radially arranged inter-grown calcite crystals; in contrast, in the haploid form many species produce holococcoliths (HOL) formed of small equidimensional calcite crystals (see Monteiro et al., 2016 and references therein). Transitions between HOL and HET life forms are now recognised as being a key component of how coccolithophore species can adapt to changing environmental conditions (Houdan et al., 2004, 2006), although the biogeography of many HOL (and HET) forms has still not been fully identified (e.g., Dimiza et al., 2008; Cros and Estrada, 2013).

Although global production of calcite by coccolithophores is still not well constrained (Bereison et al., 2007; Poulton et al., 2007), coccoliths make up the major fraction of carbonate in marine sediments (Broecker and Clark, 2009). Furthermore, extensive dissolution of coccoliths occurs both in the upper water column due to biological processes (Milliman et al., 1999; Poulton et al., 2006a) and in deep waters situated below the lysocline for calcite. Geochemical evidence suggests that low latitude calcite production is an important term in the global carbonate budget (Sarmiento et al., 2002; Bereison et al., 2007) and, in the context of climate change and ocean acidification, considerable attention is now being given to how the physiology and ecology of coccolithophores will respond to, and affect, air-sea exchanges of carbon dioxide (e.g., Bach et al., 2015; O'Brien et al., 2016). However, our knowledge of the distributions and nutritional strategies of coccolithophores across the extensive ocean gyres is still based on relatively few detailed studies (e.g., McIntyre and Be, 1967; Winter et al., 1994; Hagino et al., 2000; Haidar and Thierstein, 2001). In this paper the coccolithophore communities in equatorial and subtropical waters of the Atlantic Ocean are examined through sampling on the Atlantic Meridional Transect (AMT) programme (Robinson et al., 2006).

The growth environment of coccolithophores, and other phytoplankton, living in equatorial and subtropical waters is strongly regulated by opposing gradients in light and nutrient availability. The upper water column above the semi-permanent thermocline is characterised by low (nano-molar; <100 nmol kg$^{-1}$) inorganic nutrient concentrations and high levels of irradiance, whilst below the thermocline nutrient concentrations are relatively high (micromolar; >1 µmol kg$^{-1}$) and light levels are low. Vertical profiles of chlorophyll fluorescence show a deep chlorophyll-$a$ maximum (DCM) associated with the deep increase in nutrients (a nutricline), at depths ranging from ~60 m around the tropical Equator to as much as 150 m in the subtropical gyres. The DCM is generally regarded as neither a maximum in phytoplankton biomass or in primary production (Marañón et al., 2000; Perez et al., 2005, 2006), but marks a depth where the relative availability of surface irradiance has decreased to 1% of its surface value and where photo-acclimation results in increased cellular chlorophyll levels. Phytoplankton associated with the DCM may be considered to be light-limited rather than nutrient-limited (Venrick, 1982; Poulton et al., 2006b), whereas the situation is reversed in the upper ocean.

The phytoplankton community in the warm, stratified waters of the Atlantic are dominated in terms of abundance, biomass and primary production by small (<2 µm) picophytoplankton, mainly prokaryotes such as Prochlorococcus and Synechococcus, as well as small eukaryotes (Tarhan et al., 2006; Zubkov, 2014). The nanoplanクトon (2–20 µm), which includes most if not all species of coccolithophore as well non-califying haptophytes (Liu et al., 2009), generally contribute 10–20% of biomass (chlorophyll-$a$), with higher values in equatorial waters and higher latitude waters during spring (Marañón et al., 2001; Poulton et al., 2006b). Simple estimates of coccolithophore contributions to primary production, based on calcification rates and cellular ratios of inorganic to organic production, imply that coccolithophores account for only ~1–10% (maximum 20%) of total phytoplankton primary production in equatorial and subtropical waters (Poulton et al., 2006a, 2007). Such low coccolithophore contributions match with similar estimates made in temperate and sub-polar environments, with significant contributions (>30–40%) limited to coccolithophore blooms (Poulton et al., 2007, 2010, 2013, 2014).

Although coccolithophores are only a minor component of the phytoplankton community in the oligotrophic ocean they play an important role in the exchange of both carbon and sulphur between the atmosphere and surface waters and in the downward transport of biogenic material and its accumulation in marine sediments (e.g., Holligan et al., 1993; Malin and Steinke, 2004; de Vargas et al., 2007; Ziveri et al., 2007). It is appropriate, therefore, to ask how the growth of coccolithophores in oligotrophic waters is regulated and how it might be affected by future changes in the oceanic environment (e.g., Bach et al., 2015). Despite numerous experimental studies of photosynthesis and calcification by coccolithophores (e.g., Bach et al., 2013), as well as examination of coccoliths in sediments across glacial-interglacial transitions (Beaufort et al., 2011; Meier et al., 2014), the effects on coccolithophore ecology of future changes in ocean carbonate chemistry due to rising CO$_2$ levels remain uncertain (Bach et al., 2015). This problem stems partly from the difficulties of growing most oceanic taxa in the laboratory and an incomplete understanding of their ecology and nutritional strategies in the open ocean.

Both latitudinal biogeographical and floral depth zones have been recognised on the basis of characteristic coccolithophore species assemblages (e.g., McIntyre and Be, 1967; Okada and McIntyre, 1977; Winter et al., 1994); with further work in the Atlantic (Kinkel et al., 2000; Haidar and Thierstein, 2001; Boeckel and Baumann, 2008) and Pacific (Hagino et al., 2000; Cortes et al., 2001) confirming the generality of earlier observations. Relative to the high cell density (>1000 cells mL$^{-1}$) blooms that may occur in sub-polar waters, the reported abundances for surface subtropical waters of the Atlantic are in the range of ~1–300 coccospheres (cells) mL$^{-1}$ (the higher values being generally dominated by Emiliania huxleyi), with ~25 coccospheres mL$^{-1}$ being typical of low-latitude gyre waters (e.g., Boeckel and Baumann, 2008). Seasonal variations in abundance are most pronounced in higher latitude waters (>30°N/S) (e.g., Knappertsbusch and Brummer, 1995; Boeckel and Baumann, 2008; Baumann et al., 2008) and in equatorial upwelling regions (e.g., Kinkel et al., 2000), as a result of changes in water column stratification and nutrient distributions.

In this paper the results of SEM analyses of coccolithophore samples collected at depths of 5–300 m between 47°N and 43°S on four AMT cruises during the period 2003–2005 are reported. The AMT programme provides an excellent platform to examine coccolithophore dynamics as it provides a wealth of ancillary environmental and ecological information to provide valuable contextual perspectives to view coccolithophore ecology in the wide expanses of the oligotrophic Atlantic Ocean. Our analysis of the coccolithophore database resulting from SEM analyses has four main aims: (1) to describe the general species composition of equatorial and subtropical communities in terms of species composition, relative abundance and diversity; (2) to examine the variability in vertical structure of the community in relation to light and nutrient availability; (3) to compare and contrast vertical biogeography with latitudinal variability in species composition; and (4) to investigate whether any temporal (inter-cruise) variability in
species composition is evident across the short time period of sampling (2003–2005).

2. Materials and methods

2.1. Sampling

The cruise tracks and dates for the cruises on which samples were collected for analyses of the coccolithophore communities are shown in Fig. 1. Coccolithophore data are analysed in relation to several hydrographic provinces across the Atlantic Ocean: Northern Gyre waters (NG, 35°N–10°N), equatorial waters (EQ, 10°N–10°S), and Southern Gyre waters (SG, 10°S–30°S), with temperate (TMP) waters >35°N and >30°S (see Robinson et al., 2006; Poulton et al., 2006b; Fig. 1). Seawater samples were collected during daily pre-dawn (02:00–04:00 h, local time) and mid-morning (11:00–12:00 h) deployments of a rosette sampler fitted with 24 water bottles (20 L), a Sea-Bird 9/11 CTD, and a Chelsea MKIII Aquatracka fluorometer. Chlorophyll-a and nitrate concentrations were measured at 10–15 depths from each CTD cast, whilst samples for coccolithophore enumeration were collected from 5 to 13 water depths over the upper 300 m. The standard sampling depths for the AMT programme during 2003–2005 (Poulton et al., 2006b; Robinson et al., 2006) were the depths of 55, 33, 14, 1 and 0.1% of surface irradiance (with optical depths of 0.6, 1.1, 2.0, 4.6 and 6.9, respectively); additional samples were collected at selected stations during AMT 14 to provide a more complete description of properties across the DCM down to 300 m.

Light depths were determined from the previous days light measurements, or assuming that the deep fluorescence maximum (DCM) approximates the depth (Z_{DCM}; m) of 1% surface irradiance (optical depth = 4.6). The vertical attenuation coefficient of Photosynthetically Active Radiation (PAR; m⁻¹) was determined for each station as 4.6/Z_{DCM} and subsequent light depths determined as optical depth divided by the vertical attenuation coefficient. Optical depths for samples not collected at the five standard light depths were calculated as depth (Z; m) x vertical attenuation coefficient (m⁻¹). Sea-surface temperature was determined from the CTD sensors during daily deployments. Chlorophyll-a measurements were made on 250 mL water samples filtered onto Whatman GF/F filters, extracted in 90% acetone for 18–20 h at 4°C, and measured on a TD700 Turner Designs fluorometer calibrated with a pure chlorophyll-a standard (Sigma-Aldrich, UK) (see Poulton et al., 2006a, 2006b for further details). Micromolar (µmol kg⁻¹) concentrations of nitrate were measured with a 5-channel Technicon, segmented flow colourimetric auto-analyser (Bran + Luebbe AAII) (see Poulton et al., 2006a, 2006b for further details).

2.2. Coccolithophore enumeration and identification

For coccolithophore enumeration, 1–2 L of seawater was taken from 20 L Niskin bottles mounted on a CTD carousel, and filtered under gentle suction onto 25 mm 0.45 µm polycarbonate filters which were then dried at room temperature and stored in sealed Perspex boxes. The latitudes and depths for which samples were collected on the AMT 12 (May 2003) and AMT 14 (May 2004)
counts of Rhabdophora xiphos are likely to have been underestimated due to the occurrence of aggregates of coccospheres on the SEM filters; also this species could not always be clearly separated from Palusphaera vaneldii. The coccoliths of some species are readily detached on filters prepared for SEM analysis and in such cases aggregates of identical coccoliths were counted as a coccosphere. The full database of counts used in this study is available from the authors and via the British Oceanographic Data Centre (www.bodc.ac.uk). A total of 171 coccolithophore taxa (Appendix A) were identified in the 199 samples examined from the four AMT cruises. These taxa include recognised alternate life history phases (HET and HOL), three unnamed species (Young et al., 2003) and a few life history combinations for which two species names are still used (see Cros and Fortuno, 2002).

The low cell numbers and high diversity characteristics of subtropical coccolithophore communities (i.e., <30 cells mL$^{-1}$) lead to concern over the accuracy of SEM analyses on an examination volume (15 mL) which is a relatively small fraction of the filtered volume (1–2 L) to fully quantify the cell abundance and species diversity. One method to address such issues is to investigate how similar repeat counts of a single sample or of replicated samples are of one another in terms of cell counts and species composition. In this study, four replicate counts were performed on representative samples from below (120 m, 1% surface irradiance; optical depth = 4.6) and above (31 m, 55% surface irradiance; optical depth = 0.6) the DCM (Table 1; Fig. 3).

In general, there was good reproducibility of total numbers of taxa (species richness) and of relatively abundant cells (i.e., >1 mL$^{-1}$). However, for both sampling depths, many taxa were recorded in only one or two of the replicates, although these made a relatively small contribution to total cell counts; for the deep sample 60% of such taxa comprised only 31% of the total cell count.
Table 1

<table>
<thead>
<tr>
<th>AMT 12, CTD-26, 120 m</th>
<th>AMT 14, CTD-71, 31 m</th>
</tr>
</thead>
<tbody>
<tr>
<td>Count volume (mL)</td>
<td>No. of taxa</td>
</tr>
<tr>
<td>Stub 1</td>
<td>15.1</td>
</tr>
<tr>
<td>Stub 2</td>
<td>15.1</td>
</tr>
<tr>
<td>Stub 3</td>
<td>15.0</td>
</tr>
<tr>
<td>Stub 4</td>
<td>15.0</td>
</tr>
<tr>
<td>Cumulative</td>
<td>60.2</td>
</tr>
<tr>
<td>Mean (±S.D.)</td>
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</tbody>
</table>

Fig. 3. Histograms of cumulative species richness in selected samples from AMT 14 (CTD 71, 31 m; (a, b) and AMT 12 (CTD 26, 120 m; c). Replicate counts from the same Scanning Electron Microscopy stub (a) and repeat counts from the same sample but different stubs (b, c) are shown. The cumulative volume (mL) counted for each sample is given as a value above each histogram. Horizontal bars on (b) and (c) indicate the number of species identified in each sample.

for and surface the corresponding figures were 67% and 18%. The numbers of taxa recorded as a function of volume examined are shown in Fig. 3. For both types of sub-sampling it appears that a complete description of the coccolithophore flora can only be achieved by examining relatively large (>100 mL) volumes of seawater. Thus SEM counts based on small seawater volumes (e.g., <15 mL as used for this study) give a measure of coccolithophore species richness whilst the abundance of rarer coccolithophore taxa can be described in relative terms only. To overcome such issues it is necessary to transform and pre-treat the resulting species count data (i.e., percentage abundances), and a logarithmic transformation (log(X + 1)), which will all remove the influence of low abundance species and focus the analysis on compositional changes rather than numerical changes (Clark and Warwick, 2001).

2.3. Statistical analysis

Multivariate statistical techniques were used to assess the vertical and horizontal (dis)similarity in species composition of samples, as well as to examine whether any inter-cruise differences in species composition existed, using PRIMER-6 (v 6.1.6, PRIMER-E Ltd.) (Clark and Warwick, 2001). Bray-Curtis Similarity was calculated from standardised count data, with the exclusion of certain taxa considered to be in such low occurrences as to be rare (herein defined as consistently contributing <5% to total cell numbers), and with a Log(X + 1) transformation. Cluster analysis and non-metric Multi-Dimensional Scaling (nMDS) ordinations were then performed, with related ANOSIM (Analysis of Similarity) and SIMPER (Similarity Percentages) performed on the resulting patterns, where appropriate (Clark and Warwick, 2001). ANOSIM allows statistical comparison of the variation in species abundance and composition between groups of samples. SIMPER allows statistical identification of which species are primarily responsible for differences between groups of samples, with ANOSIM assessing the significance of the difference between groups. PRIMER was also used to calculate Pielou’s Evenness (J) which is a measure of the spread of cell abundances between the species present (Clark and Warwick, 2001).

The approach to the statistical examination of vertical, latitudinal and temporal patterns in species composition followed in this study follows a logical order of starting the analysis on a small set of samples (or species) and expanding our interpretation and analysis on this dataset by including other parts of the AMT database in steps. The main analysis focuses initially on the common (>95% total cell numbers) taxa, although an analysis of the rare species in the same manner to identify any potential patterns is also performed. The analysis follows the order: (a) firstly an examination of three high-resolution profiles from AMT 14 to identify key floral groups and vertical patterns in species composition (Section 3.1); (b) this vertical analysis is then widened to include the full cruise datasets from AMT 12 and AMT 14 (Section 3.2); (c) then consideration of latitudinal patterns in species composition based on just AMT 14 surface samples to identify key floral groups and patterns in species composition (Section 3.3); (d) this latitudinal analysis is then widened to include the full database (AMTs 12, 14, 15 and 17) to identify any inter-cruise differences or similarities in surface waters (Section 3.4); and (e) finally, an analysis of the rare species distribution, in terms of both vertical and latitudinal patterns (Section 3.5), using similar methods as used in the previous sections. The focus on AMT 14 is driven by this cruise representing the best sampled (vertically and laterally) AMT cruise, whilst only surface samples are included in the AMT database for AMT 15 and AMT 17.

3. Results

3.1. Vertical community structure: AMT 14 analysis

An in-depth analysis of vertical variability in coccolithophore species composition is firstly carried out using three high-resolution profiles from the AMT 14 cruise. These three stations represent the southern gyre (SG), equatorial waters (EQ) and northern gyre (NG) regions of the Atlantic (Figs. 1, 4 and 5). At each of these stations the numbers of taxa and cells were highest in the surface layer above the depth of the DCM (1% surface irradiance;
optical depth = 4.6), and then declined to minimum values below the depth of 0.1% surface irradiance (optical depth = 6.9) (Fig. 4a).

The relative abundance of cells and taxa within the DCM appears to vary between the two gyres, which may potentially be linked to seasonality, both being relatively low in the autumn (SG) when surface light is declining compared to the spring (NG) when surface light is increasing. In the EQ region, where the DCM and nutricline are shallow compared to the subtropical gyres (e.g., Fig. 2 for AMT 14), the decline in numbers of cells and taxa with depth are less marked. All three profiles show a relatively greater drop in cell number than in species richness below the DCM (Fig. 4a).

Each of the three profiles from AMT 14 (Fig. 4b) were characterised by a change in community structure of the coccolithophore population with depth as indicated by the fall in Bray-Curtis Similarity to <50% (with reference to the surface sample at each site) at or slightly above the depth of the DCM. In contrast, $J_0$ was relatively constant down to the DCM (Fig. 4b); since numbers of taxa for each profile were fairly uniform above the DCM it appears that changes in species composition with depth in the surface layer were not accompanied by changes in the relative abundance of the dominant species. In the two subtropical gyres the sharp drop in $J_0$ below the DCM suggests that the coccolithophore population below the depth of 1% surface irradiance (optical depth = 4.6) was distinct from that in surface waters in terms of both species composition and dominance by fewer species (Fig. 4a). However, $J_0$ increased again below the depth of 0.1% surface irradiance (at

Fig. 4. Vertical profiles of coccolithophore community characteristics against optical and percentage surface irradiance depths for three AMT 14 stations taken to be representative of the Southern Gyre, equatorial waters, and the Northern Gyre. (a) Normalised chlorophyll a (Chl), coccosphere abundance (cells mL$^{-1}$), and the number of coccolithophore taxa (species richness) (S). (b) Normalised chlorophyll a (Chl), Pielou’s Evenness ($J_0$), and Bray-Curtis Similarity ($Sim$) referenced to the near-surface sample. The depth (m) of indicative levels of percentage surface irradiance are given against the horizontal dashed lines indicating the 10%, 1% and 0.1% surface irradiance depths.

optical depths >8) and absolute depths deeper than 200 m, although these depths were characterised by very low cell numbers. The depth distribution of the abundant coccolithophore taxa (i.e., >95% of total numbers) for the three stations in Fig. 4 (AMT 14) was further examined by plotting depth profiles of each species’ cell abundance normalised to its maximum cell abundance observed in that profile (plots not shown). In our study, normalised abundance is the abundance in each sample relative to the maximum observed in the dataset; so a value of 0 would represent zero abundance and 1 would represent that species’ maximum abundance. A species’ depth habitat was then classified as the uppermost depth horizon where the maximum value (i.e., 1) and consistent values >0.75 were observed between the main relative surface irradiance depths presented in Fig. 4. Depth habitats were compared across the three stations and where, in a few cases, species had high normalised abundances in more than one depth zone, the overall depth habitat was defined as the one common across all stations. This analysis of the depth distribution of the more abundant coccolithophore taxa recorded at the three stations identified three floral groups in our study (Table 2) which are taken to characterise the Upper Euphotic Zone (UEZ, >10% surface irradiance), Lower Euphotic Zone (LEZ, 1–10% surface irradiance), and Sub-Euphotic Zone (SEZ, <1% surface irradiance). As the DCM is generally considered to mark the base of the euphotic zone (i.e., 1% of surface irradiance), the deepest coccolithophore zone is referred to as the ‘sub-euphotic’ rather than ‘lower photic’ as in other studies (e.g., Winter et al., 1994).

The vertical distributions of each of these floral groups at the three AMT 14 stations analysed, and of the rare species...
Samples in each depth zone (across the three stations) in which species were analysed was assigned to the photic zone of overall maximum abundance. The number (n) of the UEZ, LEZ and SEZ groups (see Table 2) at the SG station.

In the Sub-euphotic Zone (SEZ), species below this depth at the EQ and NG stations reflects relatively similarity = 55.2%; Table 3). The same pair-wise analysis for all UEZ and LEZ are relatively weaker (R-statistic 0.19, p < 0.05, dissimilarity = 70.7%), whereas the differences between UEZ and SEZ (R-statistic 0.97, p < 0.001, average Bray-Curtis dissimilarity = 79.1%) and

 Differences in species composition between UEZ and SEZ (R-statistic >0.80) in terms of species composition and significant (p < 0.001) R-statistics from the ANOSIM analyses. For both sets of analyses (selected AMT 14 stations and all AMT stations), the strongest statistical differences in terms of species composition were always seen between the UEZ and SEZ (R-Statistics > 0.9, p < 0.001) with average Bray-Curtis dissimilarity levels of almost 80%.

3.2. Vertical community structure: comparison between AMT 12 and 14

Vertical segregation of coccolithophore taxa during AMT 14 is clearly supported by analysis of normalised abundance (Figs. 4 and 5, Table 2) and multivariate comparison of species composition between the different sample groupings (Table 3). To test how widely these patterns exist in the fuller AMT database a multivariate statistical analysis of species composition in all AMT 12 and AMT 14 samples was performed. Coccolithophore species structure for all AMT 12 and AMT 14 samples (total 150) was investigated through calculation of Bray-Curtis Similarity and then ordination by non-metric Multi-Dimensional Scaling (nMDS). nMDS ordinations showing sample distribution with respect to cruise number, hydrographic region and optical depth are shown in Fig. 6a–c, respectively.

The nMDS ordination (Fig. 6) shows several interesting patterns in species composition, with the relative distance between samples indicative of their relative (dis)similarity. Samples with statistically similar species composition are closely aligned, whereas samples with low statistical similarity in terms of species composition are more widely spaced. Hence, the overlap in the distribution of samples from the two cruises (AMT 12 and 14) suggests strong similarities in species composition, especially for the gyre and equatorial regions (Fig. 6a and b). In terms of hydrographic region, the temperate (TMP) samples, mainly from shallow water depths, appear as a variable but relatively distinct group (Fig. 6b). In terms of irradiance levels, expressed as optical depth, the deep samples (from the depths of the 0.1% and, to a lesser extent, the 1% surface irradiance) appear as a variable but relatively distinct group (Fig. 6b). In terms of irradiance levels, expressed as optical depth, the deep samples (from the depths of the 0.1% and, to a lesser extent, the 1% surface irradiance) appear as a variable but relatively distinct group (Fig. 6b). In terms of irradiance levels, expressed as optical depth, the deep samples (from the depths of the 0.1% and, to a lesser extent, the 1% surface irradiance) appear as a variable but relatively distinct group (Fig. 6b). In terms of irradiance levels, expressed as optical depth, the deep samples (from the depths of the 0.1% and, to a lesser extent, the 1% surface irradiance) appear as a variable but relatively distinct group (Fig. 6b). In terms of irradiance levels, expressed as optical depth, the deep samples (from the depths of the 0.1% and, to a lesser extent, the 1% surface irradiance) appear as a variable but relatively distinct group (Fig. 6b).
Fig. 6. Normalised Multi-Dimensional Scaling (nMDS) ordination and bubble plots showing latitudinal and depth variability in coccolithophore community structure for AMT 12 and AMT 14: (a) by cruise; (b) by latitudinal (TMP, Temperate waters; EQ, equatorial waters; NG, Northern Gyre waters; SG, Southern Gyre waters); (c) by optical depth; (d) by coccosphere abundance (log); (e) for UEZ taxa (relative abundance to other floral groups); (f) for LEZ taxa (relative abundance to other floral groups); (g) for SEZ taxa (relative abundance to other floral groups); and (h) for rare taxa (relative abundance to other floral groups). The samples from the three high-resolution depth profiles presented in Figs. 4 and 5 are identified in panel (a) as filled square symbols. Stress on the two-dimensional plot (see (a)) was 0.17 which indicates a 'good representation of the data in two-dimensional space' (Clark and Warwick, 2001).
and 0.1% surface irradiance levels, respectively) the mean cell counts were 24.4, 29.3 and 12.9 mL⁻¹, respectively.

The relative cell counts for the different floral groups identified in Table 2 are shown in Fig. 6e-h (i.e., the relative abundance of taxa from each floral group in each sample to total numbers) for the two AMT cruises. The upper optical depths (0.6-2.0; 55-14% surface irradiance; Fig. 6c) of the gyre (NG, SG) and equatorial (EQ) regions (Fig. 6b) are characterised by abundant UEZ taxa (Fig. 6e), and the lowest optical depth (6.9; 1% surface irradiance; Fig. 6c) of the same regions by SEZ taxa (Fig. 6g). In contrast, LEZ taxa are relatively widely distributed (Fig. 6f), extending into the temperate region (Fig. 6b) and from the surface down to as deep as the 1% surface irradiance level (optical depth = 6.9; Fig. 6c). Rare taxa (Fig. 6h) are relatively evenly distributed in terms of latitude (Fig. 6b) and depth (Fig. 6c), with maximum cell numbers in the UEZ (Fig. 6e) and temperate waters (Fig. 6b); however, they are not a homogenous group, with different species occurring at different depths and latitudes (data not shown) as has been found for the common species (see Section 3.1).

3.3. Horizontal (meridional) community structure: AMT 14 analysis

Based on routine sampling from 5 depths per station it is not possible to analyse objectively latitudinal variations in the LEZ and SEZ coccolithophore flora as samples collected at the depths of 1% and 0.1% surface irradiance might, or might not, correspond to the maximum cell abundances of characteristic species for these depths (see Fig. 5b). On the other hand the surface samples (55% of surface irradiance) may be considered as representative of the UEZ flora (see Fig. 5) and hence our horizontal (meridional) analysis of species composition focuses on only surface samples. Again, AMT 14 is chosen as the first cruise to base the analysis on, with the next section (Section 3.4) widening the analysis to include all four AMT cruises.

Results of a compositional statistical analysis of AMT 14 surface samples (55% relative surface irradiance only) are shown in Fig. 7. Generally, coccolithophore cell numbers in surface waters between 30°S and 35°N ranged from 20 to 40 cells mL⁻¹, and the number of taxa (species richness) in each sample ranged from 25 to 45 (Fig. 7a). The highest coccosphere numbers were recorded from temperate stations (>30°S) and the lowest from equatorial stations. The number of taxa were relatively high (>30) within surface waters of the subtropical gyres (NG, SG), and decreased at the northern temperate station and at some equatorial and Northern Gyre stations; for example, the surface (13 m) sample from the AMT 14 14.8°N station gave a coccosphere count of just 2.2 mL⁻¹ for just 10 taxa (Fig. 7a).

Species composition in surface samples across the full AMT 14 cruise was examined by comparing statistically one station in the NG (22.3°N) with all the other surface samples in terms of Bray-Curtis Similarity. This comparison showed that the coccolithophore communities were broadly comparable (i.e., ~50% similarity) in terms of species composition across a broad latitudinal range from 35°N to 32°S (Fig. 7b). A plot of Pielou’s Evenness (Fig. 7b) for surface samples also suggests that the surface coccolithophore communities were comparable in terms of species diversity and dominance from 30°N to 30°S, except at higher latitudes where high cell densities (typically 10-100 cells mL⁻¹) of LEZ taxa such as Emiliania huxleyi, Gephyrocapsa spp., and Calcidiscus leptoporus were found. However, as shown in Fig. 7c, the relative abundance of individual taxa (e.g., E. huxleyi, Umbellosphaera tenus and U. irregularis) between stations over 20 km apart was highly variable and latitudinal peaks in relative abundance were not always consecutive.

Using a similar approach for assessing the horizontal (latitudinal) distribution of coccolithophore species as used in Section 3.1 for vertical variability, the normalised species distribution for all species which represented over 5% of total numbers in surface samples was plotted again in order to determine whether each species’ maximum relative abundance was consistently found (>0.75) within a specific hydrographic province (plots not shown). The results from this analysis are presented in Table 4, alongside the vertical floral group that each taxa (where appropriate) is aligned with. The nine UEZ taxa (see Table 2) are found mainly in the subtropical gyres (NG, SG), whilst LEZ taxa are found in both equatorial (EQ) and Temperate waters (Table 4). Of the SEZ taxa listed in Table 2, three species are found in surface temperate waters (Algirosphaera robusta, Calcosolenia murrayi, Florisphaera profunda), whereas the other four SEZ species were never observed in significant numbers in surface waters during AMT 14 (i.e., they were consistently <5% of total numbers and eliminated from the analysis). The relative abundances of these three biogeographic floras are plotted in Fig. 7d, along with the distribution of the rare (<5% of total numbers) surface water taxa. As expected, the gyre, equatorial and temperate florae reach maximum relative abundances in each of these regions, whilst the rare flora are found throughout the transect, apart from in temperate waters at either end of the transect (Fig. 7d).

A comparison of the Bray-Curtis Similarity between successive vertical samples at each station representing the UEZ, LEZ and SEZ (i.e., the statistical similarity between the 55% and 14%, 55% and 1%, 55% and 0.1% relative surface irradiance depths, respectively) again shows clear vertical gradients in species composition (Fig. 7e; see also Fig. 4b). Temperate stations are characterised by weak vertical gradients in Bray-Curtis Similarity, whilst relatively stronger vertical declines in similarity can be seen in both subtropical gyre and equatorial waters. Also, there is a marked contrast between the SG and NG samples, with high similarity (>70%) between the 55% and 14% species composition in the north which declines to ~50% in the south. A comparison of Fig. 7b and e shows that vertical gradients in species composition (in terms of Bray-Curtis Similarity) are much stronger than horizontal ones; similarity remains around 50% when comparing a sample from the NG with all samples from 30°N to 32°S (Fig. 7b), whereas similarity drops sharply below 50% when examining trends vertically over the upper 200 m (Fig. 7e). These trends highlight how there is sharper dissimilarity in coccolithophore species composition vertically over a few tens of metres than there is horizontally, over hundreds to thousands of kilometres.

3.4. Comparison between different cruises for surface communities

Analysis of meridional patterns of species composition in surface samples from AMT 14 shows several distinct features and trends in the level of statistical similarity between communities in different hydrographic provinces (Fig. 7). In this section this meridional analysis is widened to include UEZ samples (55% surface irradiance) from all four AMT cruises (AMTs 12, 14, 15 and 17), again using comparisons of Bray-Curtis Similarity in species composition in nMDS ordination (Fig. 8). In relation to cruise number, the northern spring/southern autumn (AMTs 12 and 14) and the northern autumn/southern spring (AMTs 15 and 17) samples appear to form distinct groups (Fig. 8a), a difference that may be attributable to seasonal changes in community composition rather than cellular abundances (due to the standardisation of the counts and pre-treatment of the data). When plotted in relation to hydrographic province the temperate samples form the most distinctive group and are the most scattered (Fig. 8b), again suggesting a potential seasonal signal. Variability between cruises is also evident in the equatorial and gyre waters of both hemispheres as samples from each temporal cruise pairing (i.e., AMT 12 and 14 versus AMT 15 and 17; Fig. 8a) are clearly distinct in the nMDS.
Hence, the species composition of cruises sampled in the same season but different years appears to be more similar than between successive cruises in different years; i.e., implying that seasonal differences in coccolithophore species composition are stronger than inter-annual differences. To further test the statistical significance of the sample segregation shown in Fig. 8 an identical analysis used on spatial trends in species composition (Fig. 6) is applied to the dataset. Table 5 shows the ANOSIM and SIMPER analyses on the data presented in Fig. 8, whilst species level variability is further examined (Table 6) in the context of changes in sea-surface temperature and chlorophyll-α concentration. The ANOSIM analysis supports the trends identified, with statistically significant (p < 0.05) differences in surface coccolithophore communities between AMT 12 and AMT 14 (May) versus AMT 15 and AMT 17 (September–October) for all four biogeographic provinces. Although the highest dissimilarity between seasonal samples was found for Temperate waters (69.1%), the other regions still had dissimilarity >55% (57.5–62.6%) and statistically significant (p < 0.01) R-statistics (Table 5). These results support the perspective that seasonal differences in coccolithophore species composition are stronger than inter-annual differences and the multivariate statistics are identifying seasonal variability (within the limited time-series of sampling).

Examining the raw count data further elucidates the compositional changes identified in Fig. 8, although it should be noted that due to the data treatment (standardisation, log(X + 1)) in surface coccolithophore communities between AMT 12 and AMT 14 (May) versus AMT 15 and AMT 17 (September–October) for all four biogeographic provinces. Although the highest dissimilarity between seasonal samples was found for Temperate waters (69.1%), the other regions still had dissimilarity >55% (57.5–62.6%) and statistically significant (p < 0.01) R-statistics (Table 5). These results support the perspective that seasonal differences in coccolithophore species composition are stronger than inter-annual differences and the multivariate statistics are identifying seasonal variability (within the limited time-series of sampling). Examining the raw count data further elucidates the compositional changes identified in Fig. 8, although it should be noted that due to the data treatment (standardisation, log(X + 1))

Fig. 7. Latitudinal trends of coccolithophore composition in samples from the near-surface (optical depth = 0.6 (55% surface irradiance)) during AMT 14. (a) Chlorophyll a (Chl, mg m⁻³), cell abundance (cells, mL⁻¹) and the number of coccolithophore taxa (species richness) (S). (b) Pielou's Evenness (J') and Bray-Curtis Similarity (Sim) referenced to a Northern Gyre station at 22.3°N. (c) Normalised abundance of Umbellosphaera tenuis, U. irregularis and E. huxleyi. (d) Relative abundance of coccolithophore flora (see Table 4) representative of Temperate waters, Gyre waters (northern and southern), and equatorial waters, as well as the rare group (consistently <5% of total cell numbers). (e) The degree of similarity of species composition from the 14%, 1% and 0.1% surface irradiance depths compared to the 55% sample (see a) expressed as percentage Bray-Curtis Similarity (Sim). Vertical dashed lines in panel (a) indicate the positions of the major hydrographic provinces (TMP, temperate waters; NG, Northern Gyre waters; EQ, equatorial waters; SG, Southern Gyre waters).
Table 4
Coccolithophore floral groups based on the latitudinal range of their maximum abundance relative to hydrographic provinces (see Fig. 1; TMP, Temperate waters; EQ, equatorial waters; NG, Northern Gyre waters; SG, Southern Gyre waters). The list is based on taxa found in 16 surface (55% surface irradiance) samples from AMT 14 and which represented >5% of total cell numbers in at least one surface sample. The affiliation of taxa in terms of the vertical floral zones (see Table 2) are also indicated (UEZ, Upper Euphotic Zone; LEZ, Lower Euphotic Zone; SEZ, Sub-euphotic Zone).

<table>
<thead>
<tr>
<th>Taxa</th>
<th>UEZ</th>
<th>LEZ</th>
<th>SEZ</th>
</tr>
</thead>
</table>
| Algixosphaira robusta (SEZ) | Acanthocha quattrospina | Calcosolenia sibogae | Algisphaera sp.
| Algisphaera ordinata | Anthosphaera frigida | Calcosolenia anomala | Calgisphaera sp.
| Allysphaera calcifer | Anthosphaera periporata | Calcosolenia grandis | Calgisphaera sp.
| Alysphaera pinnigera | Calyptrolithina multipora | Calcosolenia leptoporus | Calgisphaera sp.
| Alysphaera quadriradiata | Coccolithus spp. (LEZ) | Calcosolenia marina | Calgisphaera sp.
| CalciosPHAera vulgaris (SEZ) | Ceratolithus sp. (LEZ) | Calcosolenia marginata | Calgisphaera sp.
| Calcosolenia murrayi (SEZ) | Ceratolithus spp. (LEZ) | Calcosolenia magnifica | Calgisphaera sp.
| Calyptrolithina divergens (LEZ) | Cylindrosphaera sp. | Calcosolenia magdalenensis | Calcisphaera sp.
| Emiliana huxleyi (LEZ) | Discosphaera tubifera (LEZ) | Calcosolenia americana | Calcisphaera sp.
| Florisphaera profunda (SEZ) | Helicosphaera carteri | Calcosolenia carteri | Calcisphaera sp.
| Gephyrocapsa ericsonii (LEZ) | Helicosphaera carteri | Calcosolenia costata | Calcisphaera sp.
| Gephyrocapsa muelleriae (LEZ) | HELIOSphaera carteri | Calcosolenia intermedia | Calcisphaera sp.
| Gephyrocapsa ornata (LEZ) | Papposphaera rotula | Calcosolenia marginata | Calcisphaera sp.
| Helicosphaera pacimomentum | Poricalyptra aureiscina | Calcosolenia hollandeza | Calcisphaera sp.
| Oolithotus antillarum (LEZ) | Rhabdosphaera clavigera (LEZ) | Calcisphaera magnifica | Calcisphaera sp.
| Oolithotus fragilis (LEZ) | Rhabdosphaera xiphos (LEZ) | Calcisphaera magnifica | Calcisphaera sp.
| Ophistion spp. (LEZ) | Sisyphon sp. (LEZ) | Calcisphaera magnifica | Calcisphaera sp.
| Reticulofenestra carvalhoi | Sphaerocalyptra sp. | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera anthos | Syracosphaera anthes | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera athos (LEZ) | Syracosphaera pinchotii | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera carolus (UEZ) | Syracosphaera bantockii | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera delicata (UEZ) | Syracosphaera lena (LEZ) | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera halldalli | Syracosphaera noriitica | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera integrifera | Syracosphaera prolifera | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera m laschii | Syracosphaera pulchra | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera nana (HOL) | Syracosphaera pulchra | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera nodosa | Syracosphaera hollandeza | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera orthos | Syracosphaera reniformis (LEZ) | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera proutudens | Umbellosphaera irregularis (UEZ) | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera squamosa | Umbellosphaera tenuis (UEZ) | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera sp. (LEZ) | Umbilicosphaera hollandeza | Calcisphaera magnifica | Calcisphaera sp.
| Syracosphaera sibogae | Unknown holococcolith | Calcisphaera magnifica | Calcisphaera sp.

Table 5
Analysis of inter-cruise differences between surface coccolithophore communities during cruises in May (M) (AMT 12, n = 15; AMT 14, n = 17) and September (S) (AMT 15, n = 6; AMT 17, n = 21) for the four hydrographic provinces (see Fig. 1; TMP, Temperate waters; EQ, equatorial waters; NG, Northern Gyre waters; SG, Southern Gyre waters). ANOSIM is a one way analysis of similarities; SIMPER is a one way analysis on (dis)similarity percentages species contributions to the statistical differences between communities. The number of samples (n) in each province in each season is given in parenthesis.

<table>
<thead>
<tr>
<th>Groups</th>
<th>ANOSIM R-Statistic</th>
<th>p level</th>
<th>SIMPER Average Dissimilarity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>M-TMP (n = 9) v S-TMP (n = 8)</td>
<td>0.44</td>
<td>p &lt; 0.001</td>
<td>69.1</td>
</tr>
<tr>
<td>M-SG (n = 7) v S-SG (n = 11)</td>
<td>0.54</td>
<td>p &lt; 0.001</td>
<td>57.5</td>
</tr>
<tr>
<td>M-EQ (n = 8) v S-EQ (n = 5)</td>
<td>0.41</td>
<td>p = 0.01</td>
<td>57.6</td>
</tr>
<tr>
<td>M-NG (n = 10) v S-NG (n = 4)</td>
<td>0.53</td>
<td>p = 0.01</td>
<td>62.6</td>
</tr>
</tbody>
</table>

Comparison of mean values (± Standard Deviation) of surface coccolithophore community composition, sea-surface temperature (SST) and chlorophyll a concentrations. Hydrographic provinces (see Fig. 1) are: TMP, Temperate waters; NG, Northern Gyre waters; SG, Southern Gyre waters; EQ, Equatorial waters. Species were selected from one-way SIMPER analysis of species contributing to the upper 15% of dissimilarity between flora in different hydrographic provinces grouped by season. The number of samples (n) included in the analysis in each season and each province are also indicated.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Northern spring (AMT 12 and 14, n = 35)</th>
<th>Southern spring (AMT 15 and 17, n = 27)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>TMP (n = 9)</td>
<td>NG (n = 10)</td>
</tr>
<tr>
<td>Total cells (cells mL⁻¹)</td>
<td>51.2 (52.4)</td>
<td>24.6 (11.5)</td>
</tr>
<tr>
<td>Species richness</td>
<td>30.6 (14.4)</td>
<td>38.9 (11.1)</td>
</tr>
<tr>
<td>Pielou’s Evenness (J’)</td>
<td>0.6 (0.2)</td>
<td>0.8 (0.1)</td>
</tr>
<tr>
<td>Shannon-Weiner diversity (H’)</td>
<td>2.1 (0.7)</td>
<td>3.0 (0.4)</td>
</tr>
<tr>
<td>Emiliania huxleyi</td>
<td>0.30 (0.74)</td>
<td>0.10 (0.06)</td>
</tr>
<tr>
<td>Gephyrocapsa ericsonii</td>
<td>0.26 (0.44)</td>
<td>0.05 (0.06)</td>
</tr>
<tr>
<td>Umbellosphaera irregularis</td>
<td>–</td>
<td>0.03 (0.04)</td>
</tr>
<tr>
<td>Umbellosphaera tenuis</td>
<td>0.05 (0.07)</td>
<td>0.10 (0.10)</td>
</tr>
<tr>
<td>Rhabdosphaera xiphos</td>
<td>0.01 (0.01)</td>
<td>0.03 (0.03)</td>
</tr>
<tr>
<td>Paludicola vandelii</td>
<td>0.06 (0.03)</td>
<td>0.11 (0.05)</td>
</tr>
<tr>
<td>SST (°C)</td>
<td>17.8 (3.9)</td>
<td>22.9 (1.5)</td>
</tr>
<tr>
<td>Chlorophyll a (mg m⁻³)</td>
<td>0.18 (0.15)</td>
<td>0.04 (0.01)</td>
</tr>
</tbody>
</table>

4. Discussion

4.1. Coccolithophores in the equatorial and subtropical Atlantic Ocean

Total coccosphere (cell) counts of 150–300 per sample were sufficient to reliably quantify the abundance of species present at concentrations of >0.5 cell mL⁻¹ (see Table 1), who represented at least 95% of total cell numbers. The diversity (species richness) of coccolithophore communities is more difficult to determine due to the presence of a high number (140) of relatively rare species defined as consistently representing <5% of total numbers. The presence of a large number of rare coccolithophore species has also been described for other subtropical waters, such as the North Pacific gyre (Cortes et al., 2001; Thierstein et al., 2004). Species richness is dependent on the volume of sample examined which, for this study, was set by the number of coccospheres counted per sample and was generally in the range of 5–16 mL; consideration of Fig. 3 indicates that 50–70% of total species in any one sample were encountered depending on the volume examined. The high species richness, low evenness characteristics of phytoplankton communities, as discussed by Thierstein et al. (2004) for coccolithophores (see also O’Brien et al., 2016) and by Cermeno et al. (2014) for mixed phytoplankton populations, are likely to strongly influence ecological responses to both small and large scale variability in environmental conditions that affect cell growth and survival.

Setting the AMT dataset into the wider context of other open-ocean coccolithophore observations shows several interesting trends. The maximum observed counts for key coccolithophore taxa from the AMT dataset (199 samples) and from repeat sampling at the Bermuda Atlantic time-series (BATS) station (217 samples; Haidar and Thierstein, 2001) and the Hawaiian Oceanographic time-series (HOT) station (183 samples; Thierstein et al., 2004) in the subtropical Atlantic and Pacific oceans respectively are shown in Table 8. For the more numerous species, values are generally higher for BATS and lower for HOT compared to those from the AMT transect, reflecting relatively strong seasonality in surface stratification and nutrient levels at the subtropical gyre margin (BATS), which is poorly resolved within the AMT dataset, and persistent oligotrophic conditions...
throughout the year nearer to the gyre centre (HOT) compared to the more varied hydrography along the AMT transect (e.g., Fig. 2).

Another detailed taxonomic study of coccolithophores in the subtropical Atlantic is the work of Boeckel and Baumann (2008) who examined around 60 samples along a transect between 8°/C176 and 45°/C176. Boeckel and Baumann (2008) did not provide coccosphere counts for individual taxa, preferring instead to express sphere counts for individual taxa, preferring instead to express their results as ‘sphere units’ in order to take account of detached sphere counts for individual taxa, preferring instead to express their results as ‘sphere units’ in order to take account of detached spheres (Boeckel and Baumann, 2008).

Table 7

<table>
<thead>
<tr>
<th>Groups</th>
<th>ANOSIM R-Statistic</th>
<th>p level</th>
<th>SIMPER Average dissimilarity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Comparison by Hydrographic province (n = 150)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TMP (n = 35) v SG (n = 38)</td>
<td>0.21</td>
<td>p &lt; 0.001</td>
<td>80.7</td>
</tr>
<tr>
<td>TMP (n = 35) v EQ (n = 34)</td>
<td>0.18</td>
<td>p &lt; 0.001</td>
<td>80.1</td>
</tr>
<tr>
<td>TMP (n = 35) v NG (n = 43)</td>
<td>0.15</td>
<td>p &lt; 0.001</td>
<td>81.6</td>
</tr>
<tr>
<td>SG (n = 38) v EQ (n = 34)</td>
<td>0.14</td>
<td>p &lt; 0.001</td>
<td>80.8</td>
</tr>
<tr>
<td>SG (n = 38) v NG (n = 43)</td>
<td>0.07</td>
<td>p &lt; 0.005</td>
<td>81.0</td>
</tr>
<tr>
<td>EQ (n = 34) v NG (n = 43)</td>
<td>0.10</td>
<td>p &lt; 0.01</td>
<td>81.8</td>
</tr>
<tr>
<td>Comparison by Euphotic Zone Floral Groups (n = 85)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>UEZ (n = 41) v LEZ (n = 19)</td>
<td>0.29</td>
<td>p &lt; 0.001</td>
<td>79.3</td>
</tr>
<tr>
<td>UEZ (n = 41) v SEZ (n = 25)</td>
<td>0.58</td>
<td>p &lt; 0.001</td>
<td>90.4</td>
</tr>
<tr>
<td>LEZ (n = 19) v SEZ (n = 25)</td>
<td>0.12</td>
<td>p &lt; 0.01</td>
<td>86.8</td>
</tr>
</tbody>
</table>

Fig. 9. Normalised Multi-Dimensional Scaling (nMDS) ordination showing relative distributions of rare coccolithophore taxa recorded for all AMT 12 and AMT 14 samples: (a) by cruise; (b) by hydrographic province (TMP, temperate waters; NG, Northern Gyre waters; EQ, equatorial waters; SG, Southern Gyre waters); and (c) by optical depth. Stress on the two-dimensional plot (see (a)) was 0.23 which indicates a ‘good representation of the data in two-dimensional space’ (Clark and Warwick, 2001).

From an ecological perspective, in terms of the response of communities to environmental variability, it is important to consider the significance of the high diversity (species richness) of coccolithophore communities in oligotrophic waters (Fig. 3). Such diversity is characteristic of many types of microbes in the ocean, including the related non-calcifying haptophytes (Liu et al., 2009), generally taking the form of a few abundant species and a large number of rare ones (Logares et al., 2014), and is thought to be fundamental to the maintenance of ecosystem function and associated biogeochemical processes (Caron and Countway, 2009). The high species richness of each of the coccolithophore floral zones (UEZ, LEZ, SEZ) in the water column (see Tables 2 and 4) supports the conclusion of Logares et al. (2014) that assemblages have fairly regular proportions of abundant and rare taxa but contrasting structural patterns across space and time. Within the context of environmental change, diversity represents a genetic reservoir of community adaptability. However, for coccolithophores or other types of microbes, it will not be possible to
detect adaptation without initial detailed taxonomic or genomic descriptions of community structure, including the rare species.

Within the limits set by the AMT sampling programme (2 cruises per year, 2003–2005) inter-cruise differences in coccolithophore community composition were evident (Fig. 8a, Table 5). Such inter-cruise differences may relate to seasonal variability, though the timeframe of sampling is too limited to directly link these to seasonality. Clear and statistically significant (Table 5) inter-cruise differences were most obvious in Temperate waters (Fig. 8b), where large variations were observed in the abundance of common taxa such as *E. huxleyi*. Statistically significant differences in species composition were not limited to Temperate waters, with seasonal differences also detected in the subtropical gyres and equatorial waters (Table 5). Finally, it should be noted that, although large scale community distributional patterns can be generally related to large scale hydrographic and climatic factors, the causes for differences in the abundance of widespread taxa such as, for example, *Umbellosphaera* spp. over relatively small spatial scales (see Fig. 7c) remain unknown. High variability in rates of primary production in the oligotrophic Atlantic (e.g., Marañón et al., 2003) has been attributed to variations in rates of localised nutrient supply (see Johnson et al., 2010), and hence it appears that the structure, composition and dynamic properties of biological communities in the subtropical gyres is much less uniform than previously thought.

### 4.2. Coccolithophore biogeography: vertical and horizontal distributions

The characteristic vertical distribution of coccolithophore taxa observed at stations representative of the subtropical gyres and equatorial waters (Fig. 5) follows closely the general pattern observed for subtropical waters from earlier studies (e.g., McIntyre and Be, 1967; Winter et al., 1994). The terminology used in this study (UEZ, LEZ, SEZ) differs from that of other studies (e.g., Winter et al., 1994; Upper Photic Zone, Middle Photic Zone, Lower Photic Zone) due to a focus on the euphotic zone which supports upper ocean primary production. As the DCM is generally considered to mark the base of the euphotic zone (i.e., 1% of surface irradiance), the deepest coccolithophore zone is referred to as the 'Sub-euphotic Zone' rather than 'Lower Photic Zone', with two further zones in the upper euphotic zone (i.e., from the depth of 1% surface irradiance to the surface). The existence of the two upper euphotic zones (UEZ, LEZ) is supported by the species' normalised abundance profiles, with certain species having peaks not in the upper portion of the water column (UEZ) but deeper and above the depth of 1% surface irradiance (e.g., *Calciothecium* spp. and *G. ericsonii*; Fig. 5b).

For stations where samples were taken from just the 5 standard light depths used for AMT primary production studies (e.g., Poulton et al., 2006a, 2006b), it is difficult to describe or compare in detail the coccolithophore communities of the three vertical zones; the DCM (1% surface irradiance) is located at the transition between the LEZ and SEZ, with each of these two lower zones represented by just one sample (1% and 0.1% surface irradiances, respectively) that may or may not correspond to the maximum abundance of the key representative species (see Fig. 5). In contrast, the UEZ is better represented by three samples (55%, 33% and 14% surface irradiances).

Similarly, the horizontal (meridional) spacing of stations at which coccolithophores were sampled on AMT cruises 12 and 14 (Fig. 2a) is too coarse to show how oceanographic boundaries (see Aiken et al., 2000) or mesoscale eddy structure might relate to coccolithophore abundance or composition (e.g., Jin et al., 2016). In general, statistically similar communities in terms of species composition were found at closely spaced stations (<20 km apart, e.g., at the Equator on AMT 14; Fig. 7c), but there were wide differences in the relative abundance of common taxa (Fig. 7c). However, comparison of Fig. 6b and c shows that, in general, coccolithophore communities differed to a much greater degree vertically in the water column (surface layer, DCM, sub-DCM) than horizontally across hydrographic provinces (gyres, equatorial waters). This is further supported through comparison of vertical profiles of (Bray-Curtis) similarity (Figs. 5a and 7e) to meridional changes in similarity with latitude (Fig. 7b).

The depth distribution of the main floral groups summarised in Table 2 is fully consistent with the conclusions of Winter et al. (1994) (see also McIntyre and Be, 1967), who for subtropical waters, identified upper, middle and lower photic zones using similar reference light levels (>10%, 1–10%, < 1% of surface irradiance, with the DCM at the 1% level). Winter et al. (1994) stated that the middle photic zone was ‘not easily distinguished by a characteristic flora’, as also shown by the NMS ordination analysis (Fig. 6c) for the AMT samples, with LEZ species (optical depth = 4.6) showing considerable overlap with both the UEZ group (optical depths 0.6, 1.1 and 2; irradiance levels 55, 33 and 14%) and SEZ species floral communities.

### Table 8

<table>
<thead>
<tr>
<th>Species</th>
<th>AMT 32°5′-30°N SEM</th>
<th>AMT 32°2′/N LM</th>
<th>HO T 22°N SEM</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Calcidiscus leptoporus</em></td>
<td>R</td>
<td>1.5 (37.0)</td>
<td>2.3</td>
</tr>
<tr>
<td><em>Discosphaera tubulosa</em></td>
<td>UEZ</td>
<td>3.7 (8.2)</td>
<td>2.3</td>
</tr>
<tr>
<td><em>Emiliania huxleyi</em></td>
<td>LEZ</td>
<td>35.4 (265.8)</td>
<td>92.7</td>
</tr>
<tr>
<td><em>Florisphaera profunda</em></td>
<td>SEZ</td>
<td>42.7</td>
<td>67.5</td>
</tr>
<tr>
<td><em>Gephyrocapsa ericsonii</em></td>
<td>LEZ</td>
<td>12.0 (224.1)</td>
<td>26.2*</td>
</tr>
<tr>
<td><em>G. oceanica</em></td>
<td>R</td>
<td>4.0</td>
<td>1.4</td>
</tr>
<tr>
<td><em>Globolithus flabellatus</em></td>
<td>SEZ</td>
<td>9.6</td>
<td>5.2</td>
</tr>
<tr>
<td><em>Helicosphaera spp.</em></td>
<td>R</td>
<td>1.2</td>
<td>1.4</td>
</tr>
<tr>
<td><em>Rhodobosphaera clavigera</em></td>
<td>LEZ</td>
<td>6.2</td>
<td>4.7</td>
</tr>
<tr>
<td><em>Syracosphaera pulchra</em></td>
<td>UEZ</td>
<td>1.8</td>
<td>0.9</td>
</tr>
<tr>
<td><em>Umbellosphaera irregularis</em></td>
<td>UEZ</td>
<td>11.2</td>
<td>16.2</td>
</tr>
<tr>
<td><em>U. tenus</em></td>
<td>UEZ</td>
<td>9.8</td>
<td>34.1</td>
</tr>
<tr>
<td><em>Umbellosphaera hubulirtiana</em></td>
<td>R</td>
<td>2.6 (8.8)</td>
<td>0.5</td>
</tr>
<tr>
<td><em>U. sibogae</em></td>
<td>R</td>
<td>1.2 (1.4)</td>
<td>0.6</td>
</tr>
</tbody>
</table>

*na* = not available.

* Counted as small coccospheres.
group (optical depths 4.6 and 6.9; 1 and 0.1%). It is not surprising therefore that some species listed for the LEZ in Table 2 have been previously assigned to the layers above (e.g., *Ceratolithus* spp.) or below (e.g., *Syracosphaera anthos*). The conclusion of Winter et al. (1994) that *Umbellosphaera tenuis*, *Syracosphaera* spp., and placolith-bearing genera (such as *E. huxleyi* and *Gephyrocapsa* spp.) show no depth preference but tend to be most abundant in the middle photic zone is generally well supported by the AMT data, although some of the diverse *Syracosphaera* group (in particular *S. pulchra*) are found mainly in the UEZ (Table 2) and *U. tenuis* is classed as a UEZ taxon that extends to greater depths than *U. irregularis* (see Fig. 5a) in the AMT analysis.

Two general features of coccolithophore vertical distribution shown by the AMT data are firstly that the Bray-Curtis Similarity index declines with respect to the surface sample throughout the UEZ and LEZ (Fig. 4b; see also Fig. 7e), whereas the total numbers of both cells and species remain uniform between the surface and the depth of 1% surface irradiance (Fig. 4). Secondly, cell numbers for some characteristic SEZ taxa (in particular *Gladio lithus flabellatus*; see Fig. 5a) reach a maximum at or below the depth of 0.1% surface irradiance (Fig. 5b). The first observation implies that despite compositional changes in species composition with depth, the relative size of the community remains consistent, and that rare taxa as well as abundant ones (Table 2) change with depth; within the AMT dataset rare taxa that were largely confined to the LEZ include *Acanthoica quattro spina*, *Alisphaera pinnigera*, *Corisphaera gracilis*, *Gephyrocapsa oceanica*, *G. ornata*, *Helicoc ophaera* spp., *Picrota murgalefi*, *Syracosphaera reniformis* and *Umbilicosphaera* spp. Peaks in species abundances well below the depth of 1% surface irradiance, where light can be assumed to be limiting to photosynthetic processes whilst nutrients are non-limiting (e.g., Poulton et al., 2006b), are likely indicative that most, if not all, SEZ species (e.g., *G. flabellatus*) have alternative nutritional strategies than autotrophy (i.e., mixotrophy or phagotrophy).

Regional anomalies in the vertical distribution of coccolithophores can be inferred from Fig. 7e. Firstly, greater similarity between samples from the depths of 14% and 0.1% of surface irradiance in temperate waters compared to subtropical waters reflects greater vertical mixing of the upper water column at higher latitudes and a more uniform environment; in autumn (i.e., at the southern stations on AMT 12 and 14), this trend can be attributed partly to populations of SEZ species, including *Florisphaera profunda*, being mixed towards the surface, whereas in spring (northern stations) UEZ and LEZ species occur throughout the water column before the DCM and associated flora becomes well established. Secondly, greater statistical similarity of species composition between samples from the 1% and 0.1% surface irradiance depths in subtropical waters of the northern hemisphere (spring) compared to those of the southern hemisphere (autumn) could be related to seasonal changes in the permanent DCM (see Letelier et al., 2004); in spring, when the depth of 1% surface irradiance deepens such that the DCM is displaced downwards into a layer where SEZ species, in particular, *F. profunda* are relatively abundant.

The UEZ and SEZ coccolithophore taxa (Table 2) are largely confined to the well-stratified upper waters (<100 m) of the equatorial and subtropical Atlantic (Fig. 7e), with their distributions extending polewards in summer months with seasonal strengthening of stratification (see Haidar and Thierstein, 2001). In contrast, LEZ taxa, and associated rare taxa (e.g., as listed above, Fig. 10b), are widely distributed with respect to both latitude and depth (Fig. 6f) and reach their maximum abundance in temperate waters (Figs. 6d and 7f) where high concentrations (>50 coccospheres mL⁻¹) of species such as *E. huxleyi* and *G. ericsonii* are observed. Hence, the LEZ taxa often represent the dominant species in temperate and high latitude waters, a phenomenon potentially linked to light availability in that light levels experienced in the LEZ (i.e., 10% of 2000 μmol photons m⁻² s⁻¹ on a 14–16 h day equates to 10.1–11.5 mol photons m⁻² d⁻¹) are similar to average mixed layer irradiances experienced by temperate coccolithophore communities (e.g., 10–15 mol photons m⁻² d⁻¹; see for e.g., Poulton et al., 2010, 2014).

4.3. Ecological and nutritional strategies of subtropical coccolithophores

A notable feature of the AMT data is a strong difference in the mean vertical distribution between HET and HOL forms of the same species (see Tables 2 and 4), with the latter consistently occurring at shallower depths as has been described for stratified waters of the Mediterranean by Dimiza et al. (2008) and Cros and Estrada (2013). The best example in the AMT dataset is provided by *Sy racosphaera anthos* (Fig. 10a); with the distribution of the HET form which belongs to the LEZ floral group often extending well below the DCM, whereas the HOL form is characteristic of the UEZ group and typically confined to surface waters. Widening this analysis to all species that have recognised HOL-HET stages in the AMT database (9 species; *Acanthoica quattro spina*, *Calidicus leptopus*, *Coccolithus pelagicus*, *Coronosphaera mediterranea*, *Helicosphaera carteri*, *Heli-
cosphaera carteri, Syracosphaera anthos, S. bannockii, S. nana, and S. pulchra) and plotting the ratio of HOL to HET cell counts (Fig. 10b) shows a clear decrease in the relative abundance of HOL forms with depth. However, there are also clearly high numbers of HET forms for some species (e.g., Acanthoica quadrotspina) in surface waters as indicated by the low HOL:HET ratios at light levels >10%.

A clear trend in the ecology of subtropical coccolithophores, in terms of floral groups, rare species and HOL-HET species combinations is that a relatively large number of cells and species are found deep in the water column (Tables 2 and 6: Figs. 4–6, 9 and 10). Such deep waters experience low light levels in both relative (<1% of surface irradiance) and absolute terms (<20 μmol photons m⁻² s⁻¹ based on a surface irradiance of 2000 μmol photons m⁻² s⁻¹ or <2 mol photons m⁻³ d⁻¹ on a 14–16 h day), which are likely to be limiting to autotrophic growth. The oligotrophic waters of the equatorial and subtropical Atlantic Ocean are characterised by a DCM which is situated on the nutricline (Fig. 2); autotrophs above the DCM are likely (inorganic) nutrient-limited, whilst below they are likely light-limited (e.g., Poulton et al., 2006b). Hence, most coccolithophore species face the problem of either insufficient inorganic (nitrate, phosphate) nutrients (UEZ and LEZ taxa) in the upper water column or insufficient light (SEZ taxa) at depth in order to compete with the dominant, small (<2 μm) photosynthetic picoplankton that have a high efficiency for the assimilation of nutrients and light energy (Zubkov, 2014).

The DCM is generally considered to be a pigment rather than a biomass maximum (i.e., composed of cells with a low carbon-to-chlorophyll ratio and hence high cellular pigmentation) across the oligotrophic Atlantic Ocean. Picoeukaryotes, autotrophic cells <2 μm in diameter, are relatively abundant in the DCM of the Atlantic subtropical gyres (<1000–5000 cells ml⁻¹; Tarran et al., 2006) and, in the northern gyre, their distribution has been shown to be closely associated with deep peaks in uptake rates for nitrate (Painter et al., 2014). Tarran et al. (2006) also reported that the maximum biomass of autotrophic nano-eukaryotes (2–10 μm diameter) occurred immediately above the DCM, where growth is likely to be enhanced close to the nutricline by some upward diffusion of nutrients and by nutrient regeneration associated with biological activity at the DCM.

For the coccolithophores, the DCM represents a transitional zone between the LEZ and SEZ floral groups (Fig. 5a); the number of coccospheres in the DCM, although dominated by LEZ taxa, is lower than in the UEZ except at equatorial stations (Fig. 4a), and important LEZ species often show a clear maximum in abundance above, rather than at, the DCM (Fig. 5b). Thus, there appears to be no clear relationship between the vertical distribution of coccolithophores and the position of the nutricline, suggesting that irradiance at this depth is too low to support nitrate assimilation by these relatively large (5–20 μm) cells. However, the LEZ group includes taxa (E. huxleyi, Gephyrocapsa spp.) that increase in abundance in response to nutrient enrichment as well as taxa (Calcidiscus, Discosphaera, Michaelsarsia) that do not (see Brand, 1994). The former probably have a similar nutritional status to the nano-eukaryotes mentioned above whereas the latter, together with UEZ taxa, are considered more characteristic of nutrient-depleted oceanic waters and, almost without exception, have not been grown successfully in culture. Little is known about the biology of SEZ taxa, even to the extent as to whether or not they all have functional chloroplasts and are autotrophic. At light levels <1% of surface irradiance it seems very unlikely that they could reach observed abundances (Table 8) by phototrophy (photosynthesis and reduction of inorganic nutrients) alone.

There are a number of references to possible mixotrophy by coccolithophores in connection with their survival in low-nutrient oceanic waters (Brand, 1994), the function of the haptonema (flagella) as a food gathering organelle (Kawachi and Inouye, 1995), the suggested function of modified coccoliths as ‘particle collectors’ (Aubry, 2009), and their phylogenetic linkages to heterotrophic flagellates (de Vargas et al., 2007). There is still little observational or experimental evidence for mixotrophy by coccolithophores. However, from a consideration of (i) the physiological ecology of species inhabiting oligotrophic oceanic waters (Brand, 1994), (ii) mixotrophy by non-calciifying haptophytes (Liu et al., 2009; Unrein et al., 2014), (iii) the quantitative importance of mixotrophy in oligotrophic oceanic ecosystems (Hartmann et al., 2012), (iv) the existence of heterotrophic coccolithophores in polar waters (Thomsen et al., 1991), and (v) the recognition of the significance of mixotrophy for the oceanic biological carbon pump (Mitra et al., 2014) it is highly likely that many coccolithophores are actually mixotrophic, with possible exceptions being placolith-bearing species belonging to the LEZ group which form blooms in nutrient-enriched waters (e.g., E. huxleyi, Gephyrocapsa spp., Calcidiscus spp., Umbilicosphaera spp.).

The distinct vertical profiles for the HOL and HET forms of the same species (Dimiza et al., 2008; Cros and Estrada, 2013), may therefore reflect differences in the degree or type of mixotrophy across the group; HOLs in shallower, more oligotrophic water could supplement phototrophy with heterotrophy whereas HETs, in the absence of evidence for an ability to utilise nitrate under low light at depth, could be largely or entirely heterotrophic. A shift from one life-cycle stage to another may also represent an ecological strategy (Houdan et al., 2004, 2006) in response to changes in environmental conditions. In the case of the HET form of Syracosphaera anthos (Fig. 10), nano-eukaryotes in the DCM (Painter et al., 2014) are potentially an important food source. Alternative nutritional strategies for coccolithophores in low (inorganic) nutrient conditions may also include the use of dissolved organic carbon and nutrient sources to supplement or replace photosynthetic growth, though the composition, lability and bioavailability of this material is unclear. Clearly, innovative future research on the nutritional strategies of (subtropical) coccolithophores is key to fully understanding the ecology and biogeography of such species, as well as gaining a better perspective on coccolithophore impacts on the biological carbon pump in the oligotrophic ocean.

A better understanding of the nutrition of coccolithophores will also open the way to improved interpretation of data on the distribution of coccoliths in marine sediments in relation to environmental conditions. In particular for the important palaeoindicator species Florisphaera profunda, SEZ taxa are mainly or entirely found below the DCM and if they are partly or wholly heterotrophic then their abundance is more likely to reflect the availability of living and non-living particulate material rather than of irradiance or inorganic nutrients (e.g., Molfin and McIntyre, 1990) or of a well-developed DCM (e.g., Grellard et al., 2012). Furthermore, the depths of the nutricline and the SEZ are probably not independent variables as envisaged by Molfin and McIntyre (1990), in the sense that the depth of the nutricline approximates the 1% surface irradiance level, whether it is deep or shallow, and varies with the degree of biological attenuation of light in the overlying water (and the uptake of nitrate as light becomes available; Letelier et al., 2004). Hence, a shallowing of the depth of the 1% surface irradiance will be accompanied by a shallower nutricline as nitrate uptake follows light availability.

Furthermore, oceanic ecosystems associated with a shallow thermocline are more productive, and also generally show a greater abundance of F. profunda in the water column; for example, Fig. 2 shows how the nutricline (and DCM) shallows in the equatorial region and northern gyre in spring during AMT 14 (see also Figs. 2 and 4 in Robinson et al., 2006), also at gyre margins (BATS) compared to gyre centres (HOT) (see Table 8). Therefore, it appears that changes in the relative abundance of F. profunda coccoliths in...
ocean sediments may reflect changes in the production and/or dissolution of coccoliths of UEZ and LEZ taxa. In the oligotrophic subtropical ocean with a deep nutricline, phytoplankton are relatively scarce and will be efficiently consumed by herbivores within surface waters (e.g., Poulton et al., 2006a), leading to high losses of coccoliths due to dissolution (Milliman et al., 1999), whereas at gyre boundaries and around upwelling regions, increases in coccolithophore abundance in the euphotic layer will lead to sudden downward fluxes of UEZ and LEZ coccoliths, a proportion of which may escape dissolution. However, at ocean boundaries and within enclosed seas, other factors may control the relative abundance of different types of coccoliths in the sediments (Ahagon et al., 1993); organic matter derived from continental shelf and terrestrial sources may be a significant source of food for heterotrophic coccolithophores, whilst freshwater inputs can affect the growth conditions for autotrophic taxa either positively or negatively through effects on nutrient levels, on light penetration (turbidity), and on depth of the thermocline. For example, in the western subtropical Atlantic, sediments in which G. flabellatus is more abundant than F. profunda are found in offshore regions affected by the outflow of the Amazon River, close to the continental slope affected by the Brazil Current and over the mid-Atlantic ridge (Boeckel et al., 2006); it would be of great interest to know why this happens in such contrasting situations.

4.4. Future recommendations

Though sample collection was limited to a depth resolution of only five standard light depths across much of the AMT transect, these light depths still reflected the general pattern of the vertical distribution of species composition in the equatorial and subtropical Atlantic Ocean. Ideally, higher resolution sampling (e.g., 15–20 sampling levels) would lend further support to the vertical patterns identified in this study, and if combined with increased meridional sampling frequency would also greatly expand the species level insights gained. The difficulty in reaching an examination volume which fully accounts for the incredible level of species diversity represents a limitation to gaining further insights into the biogeography and ecology of the rare species. Furthermore, the point where the cumulative species richness plateaus against examination volume (i.e., where the rarefaction curve plateaus) will likely be sample specific, also providing a limitation. However, examination of 150–300 coccospheres per sample did account for the dominant species in terms of cell numbers: with 31 out of the 171 taxa representing 95% of total cell numbers, and hence though it is not possible to fully describe the full species richness of the community it is possible to reliably analyse the vertical and meridional distribution of the numerically dominant species. In order to confidently identify seasonal and interannual variability in species composition in the subtropical ocean, a longer timeframe is required to be able to differentiate random variability from temporal trends. Such an expanded dataset, if geographically aliased in a suitable way, would also provide further insights into the biogeography of rare coccolithophore species.

The new perspectives gained from statistical exploration of coccolithophore distribution in the equatorial and subtropical Atlantic Ocean warrant further examination and confirmation. Such research will require innovative methodology and experimentation to confirm the nutritional strategies of upper-ocean and deep-dwelling coccolithophore species. For example, tracing particular uptake through the use of fluorescently labelled prey items, high-resolution microscopy to examine the presence/absence of chloroplasts, and tracing the cellular uptake of labelled compounds. Heterotrophic or mixotrophic nutrition of coccolithophores has significant implications for their biogeochemical roles; for example, the impact on dissolved inorganic carbon and alkalinity through calcification, photosynthesis, respiration and dissolution will be very different to a cell which lacks photosynthesis and has high ratios of calcification to respiration. Indeed, the cellular coupling of photosynthesis and calcification, in terms of internal (carbon) substrate competition, will be very different in a cell which attains its energetic needs through respiration rather than autotrophy. How this impacts on cellular rates of calcification and growth, and hence the fitness and competition between autotrophic and heterotrophic (partly or fully) coccolithophores is completely unknown, but a key issue to be addressed when considering the species composition of oceanic communities. Moreover, communities dominated by autotrophic, heterotrophic or mixotrophic coccolithophores will have different influences on air-sea CO$_2$ fluxes through variable roles in organic and inorganic production, and the biological carbon pump and carbonate-counter pump.

5. Conclusions

Across 199 samples collected in the equatorial and subtropical Atlantic Ocean, 171 coccolithophore taxa were identified (Appendix A); of which 140 were classified as rare, consistently representing <5% of total cell numbers in all samples, whilst 31 were observed in enough abundance to allow us to analyse their vertical and meridional distribution. From multivariate statistical analysis, strong vertical gradients in species composition were identified in equatorial and subtropical waters, which lessened in Temperate waters. Three vertical zones in terms of floral composition were identified: an upper euphotic zone (UEZ), a lower euphotic zone (LEZ), and a sub-euphotic zone (SEZ). This vertical zonation is closely tied to the light availability through the water column, and the species involved in each zone are broadly similar to those identified by Winter et al. (1994) (see also McIntyre and Be, 1967). Light levels in the SEZ are likely to be well below those required to support photosynthesis and hence it is suggested that ingestion of other plankton (mixotrophy or full phagotrophy) or dissolved organic compounds may support the growth (and hence calcification) of species at these depths.

Coccolithophore cell numbers were highest in temperate waters, whilst species richness was highest in the subtropical gyres and equatorial waters; comparison of gyre samples with other surface samples highlights relatively high similarity (>50%) across large horizontal distances, which contrasts with the sharp declines in similarity with depth seen in the gyres and equatorial waters. UEZ taxa were largely restricted to the subtropical gyres, whilst the LEZ taxa were found in both equatorial (EQ) and Temperate regions and some SEZ species were found in surface Temperate waters. LEZ species often represent the dominant species in Temperate and high latitude waters, which is potentially linked to similar light (and inorganic nutrient) climates between the LEZ in the subtropics and mixed layers in Temperate waters. Seasonal differences in coccolithophore species composition were stronger than inter-annual differences. The strongest seasonal variability seen was in Temperate waters, with elevated abundances of species such as E. huxleyi and G. ericsonii (both LEZ). Clearly the coccolithophore communities in low-latitude, low (inorganic) nutrient waters respond to temporal changes in environmental conditions and these environments are not quiescent regions of the global ocean.

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## Appendix A

List of taxa identified by Scanning Electron Microscopy. Holo-
coccolithophores (HOL) indicated in bold italics.

<table>
<thead>
<tr>
<th>Species name</th>
<th>Species name</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acanthoica maxima</td>
<td>Cyrtosphaera localae</td>
</tr>
<tr>
<td>Acanthoica quattropinna</td>
<td>Discosphaera sp.</td>
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<tr>
<td>Acanthoica quattropinna HOL</td>
<td>Discosphaera tubifera</td>
</tr>
<tr>
<td>Acanthoica sp.</td>
<td>Emlimia haluxi</td>
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<td>Algicrateria cucculata</td>
<td>Eriocystis sp.</td>
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<tr>
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<td>Florisphaera profunda</td>
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<td>Alisphaera extenta</td>
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</tr>
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</tr>
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<tr>
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<td>Gephyrocapsa sp.</td>
</tr>
<tr>
<td>Alisphaera unicorns</td>
<td>Glandolitih us flabellatus</td>
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<tr>
<td>Alisphaera sp.</td>
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<td>Coccolithus pelagicus</td>
<td>Ophistser sp.</td>
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<td>Coccolithus pelagicus HOL</td>
<td>Palusphaera vandelli</td>
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<td>Corisphaera gracilis</td>
<td>Palusphaera sp.1</td>
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<td>Corisphaera striglis</td>
<td>Pappomonas sp. Type 2</td>
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<tr>
<td>Corisphaera tyrrelliensis</td>
<td>Papposphaera lepida</td>
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**Appendix A (continued)**

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<th>Species name</th>
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<tbody>
<tr>
<td>Coronosphaera binodata</td>
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<td>Coronosphaera maxima</td>
<td>Pappomonas sp. Type 4</td>
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<tr>
<td>Coronosphaera mediterranea</td>
<td>Pappomonas sp.</td>
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<td>Papposphaera arctica</td>
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<tr>
<td>Coronosphaera mediterranea HOL wettsteinii</td>
<td>Papposphaera borealis</td>
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<td>Coronosphaera sp.</td>
<td>Papposphaera thomsoni</td>
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<td>Cyrtosphaera aculeata</td>
<td>Papposphaera sp. Type 1</td>
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</table>

**Species name**

- Syracosphaera pirus
- Syracosphaera tumlaris
- Syracosphaera hastata
- Syracosphaera didyma
- Syracosphaera castellata
- Syracosphaera reniformis
- Syracosphaera squamosa
- Syracosphaera leptolepis
- Syracosphaera sp.
- Tetralithodes quadrirlaminata
- Turrilithus latericoendites
- Umbillosphaera irregularis
- Umbillosphaera tensis
- Umbillosphaera anulus
- Umbillosphaera helburtiana
- Umbillosphaera sibogae
- Zygococcolithina amoena
- Undescribed heterococcolithohore A
- Undescribed coccolithophile
- Undescribed holococcolithophile

Appendix A (continued)

<table>
<thead>
<tr>
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<td>Syracosphaera molischii</td>
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<td>Syracosphaera nana</td>
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<td>Syracosphaera noriticola</td>
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<td>Syracosphaera ossa</td>
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References


